synthesis kit and ZAP Express cDNA synthesis kit, respectively (Stratagene). For directional cloning, 1st strand cDNA synthesis was primed with oligo-dT (the primer also contains an XhoI site) using Moloney murine leukemia virus (M-MuLV) reverse transcriptase. After 2nd strand synthesis and addition of an EcoRI adaptor, the cDNAs were digested with XhoI and size-fractionated with SEPHACRYL™ S-400 column. Fractions 4 and 5 were combined and further electrophoresed in 1% agarose gel to isolate cDNAs greater than 1 kb in size. The purified cDNAs were ligated to EcoRI/XhoI double digested Lambda ZAP II vector. The ligation product was packaged into GIGAPACK™ II Gold packaging extract (Stratagene), then plated in XL1-Blue MRF' host cells. The primary library was estimated to contain $6x10^6$ independent recombinants and has an average insert size of 1.9 kb. This oligo dT primed library was amplified once and stored in aliquots at 4°C (in chloroform) or -70°C (in DMSO). For construction of random primed library, 1st strand cDNA synthesis was primed by random hexameric oligonucleotides. The cDNAs were ligated with EcoRI adaptor, size fractionated with SEPHACRYL™ S-400 column, and cloned into EcoRI digested ZAP express vector (Stratagene). This random primed library has $6x10^6$ independent recombinants and an average insert size of 1.6 kb.--

Replace the paragraph on page 19, lines 1 to 13, with the following rewritten paragraph:

--Detection of pre-S binding proteins in labeled lysates. To reduce the levels of cellular proteins which bind to the GST component of the pre-S fusion protein or directly to the sepharose beads, labeled lysates were first preincubated twice at 4°C with a mixture of empty sepharose beads and GST-bound beads. The beads were washed extensively and used as a negative control in 8% SDS-PAGE. The precleared lysates were then incubated at 4°C for 6 to 16 hr with the specific GST-pre-S fusion protein. After extensive washing of the beads four times with lysis buffer, bound proteins were eluted from beads by heating to 95°C for 5 min, and separated on 8% SDS-PAGE under denaturing conditions. Proteins retained in the second preclearing reaction were run in parallel. The gel was fixed with 10% acetic acid, treated with AMPLIFY™ solution (Amersham), dried, and exposed. For experiments performed with the ^{125}I labeled proteins, the treatment step with AMPLIFY™ solution was omitted. Comparison of protein bands binding only to the pre-S fusion indicated the specificity of the interaction.--